

Identification of Coniferous Woods

B. FRANCIS KUKACHKA

The identification of coniferous woods is generally regarded as being more difficult than for the hardwood species. This is due to the fact that conifers are more elemental in their structure and, as a consequence, the number of diagnostic features that may be employed is proportionately smaller. Instructions are given here in the sequential use of primary diagnostic features in the identification of coniferous woods. These may be used from the data chart or by sorting marginally perforated cards. Microscopic features are necessarily emphasized because of the greater accuracy obtainable through their usage. The majority of the species described are found in the continental United States and Canada; however, several exotic species found in commercial channels are also included.

EXPERIMENTAL

Equipment

NECESSARY identification equipment includes: Suitable knives or razor blades, either for freehand or mechanical sectioning; magnifying lens of approximately 15 power; electrical hot plate or equivalent heat source; microscope capable of magnifying up to 400 or more times; a microtome if mechanical sectioning is contemplated; and microscope slides and cover glasses.

Reagents

Required reagents are: 1% solution of caustic soda, and a 50-50 solution of glycerine and 95% ethyl alcohol.

Preparation of Specimens

Because identification may be made from a piece of wood, pulp sample, or a paper sample, two different methods are used in preparing the specimens for examination.

In the case of a piece of wood, it is necessary to examine both radial and tangential sections because most of the diagnostic features will be seen from these surfaces. These sections may be prepared by cutting thin slices freehand with a razor blade or microtome knife. It is usually desirable to wet the surface prior to

cutting, or in some cases it may be desirable to boil the block for a short time in order to cut sections that are thin enough to be transparent at the higher magnifications on the microscope. The sections are then placed on a microscope slide, a few drops of the alcohol-glycerine mixture are placed over the sections, and a cover glass applied. The prepared slide is then heated to the boiling point for a minute or two to expel air from the sections. After cooling for a few minutes the slide is ready for examination.

To examine the component, cells of a pulp or paper sample, it is usually necessary to boil a small quantity of the sample for a few minutes in a 1% solution of caustic soda to remove sizing materials. The sample is then washed to remove the alkali and shaken vigorously in water to make a suspension of the cells. A few drops of the suspension are placed on a microscope slide and a cover glass is applied. It is usually not necessary to stain the fibers; however, if desired, the staining may be accomplished directly after washing the sample and before or during the shaking process.

DIAGNOSTIC FEATURES USED IN IDENTIFICATION

The present work is based primarily on features visible with a microscope and is intended primarily for wood technologists that have access to an authenticated collection of wood specimens. Detailed descriptions of the various species are not included; however, the presence of diagnostic features is shown in Table I and attention is also called to the supplementary notes on the various genera.

In the following notes, the diagnostic features selected for their value in identification are defined and their significance discussed. The majority of the anatomical features used in this key are also illustrated.

Most of the features refer to microscopic structure; however, there are also included certain general or gross features that are of value in specific instances.

It will be noted that some of the numbered features have been left blank. This was done expressly for those who may wish to enlarge the key by the addition of other species which may, in turn, necessitate the inclusion of more diagnostic features.

1. Resin Canals. When resin canals are present,

B. FRANCIS KUKACHKA, Forest Products Technologist, Forest Products Laboratory, maintained at Madison, Wis., in cooperation with the University of Wisconsin, Forest Service U. S. Department of Agriculture.

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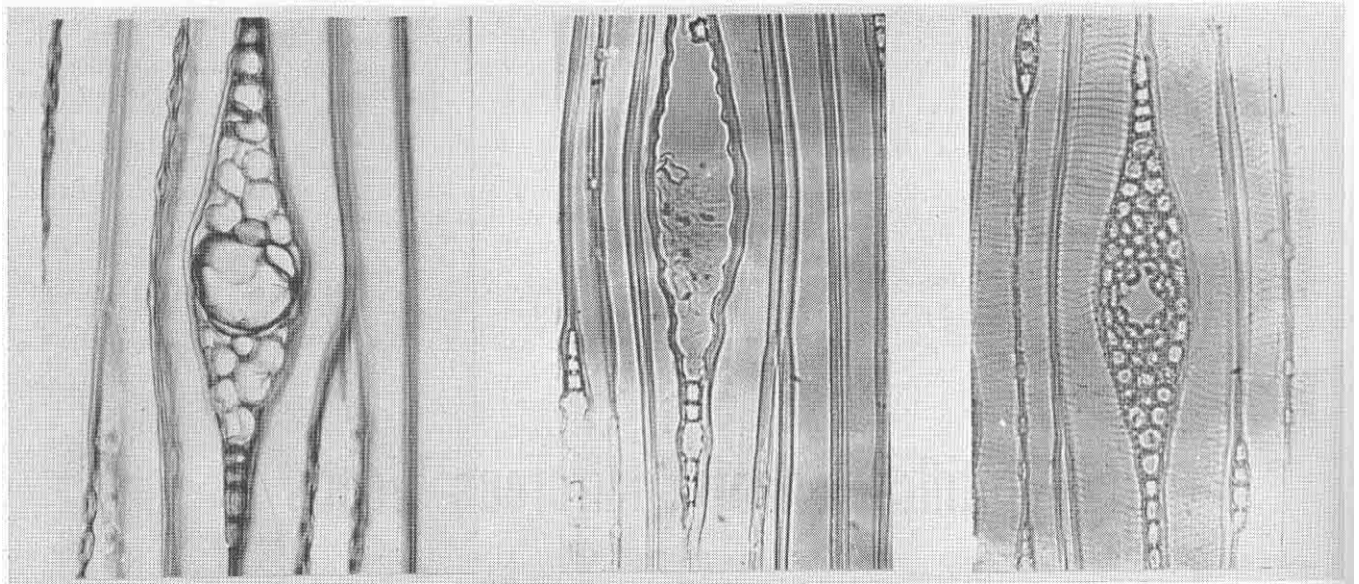


Fig. 1. Left, thin-walled epithelium and ray parenchyma intact after sectioning of embedded specimen. Center, thin-walled epithelium and ray cells torn out in specimen that was not embedded. Right, thick-walled epithelium intact after freehand sectioning of untreated specimen

they occur in both the vertical and horizontal system. On smoothly cut transverse surfaces they appear to be solitary or infrequently paired. Resin ducts vary appreciably in size and abundance and in most species can be detected with the eye. They are largest and most abundant in *Pinus* and of smaller size and fewer in number in *Picea*, *Larix*, and *Pseudotsuga*. In some instances they appear as short scratch marks on the longitudinal surfaces of the wood or may become more pronounced because of resin stains in adjacent areas. Exudation of resin from the end grain, the presence of pitch pockets, or pitch streaks are additional indicators of the presence of resin canals. The presence of normal

ducts can always be verified by an examination of tangential sections under the microscope.

2. Epithelium Cells Thick-walled. In freehand sectioning or sectioning without embedding the wood, species that possess resin canals with thin-walled epithelium cells will show the fusiform wood rays containing the canals to be devoid of an epithelium layer and, in many instances, the adjacent ray parenchyma will also be removed in the process. This condition is shown in Fig. 1 center, where the canal as well as the thin-walled parenchyma cells have been torn out in sectioning without embedding. Figure 1, left, shows a similar-type fusiform ray which had been embedded and all of the cells are intact. Figure 1, right, illustrates the thick-walled type of epithelium which has remained intact even though the section was cut freehand.

3. Blank.

4. Ray Tracheids Common. Ray tracheids may be found in many species if a careful search is made, but their rare occurrence in such cases suffices to distinguish those in which they are immediately seen. In certain genera they may be confined to the margins of the wood rays, as in *Tsuga* and *Larix*, while in others, for instance *Pinus*, they may be both marginal and interspersed. When the ray tracheids are nondentate they may be

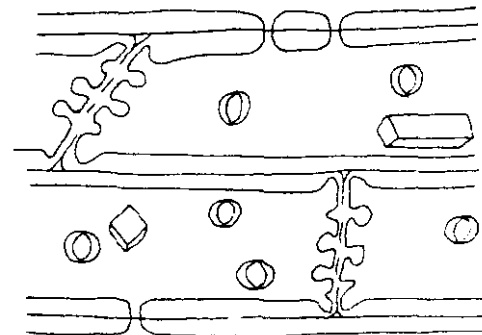
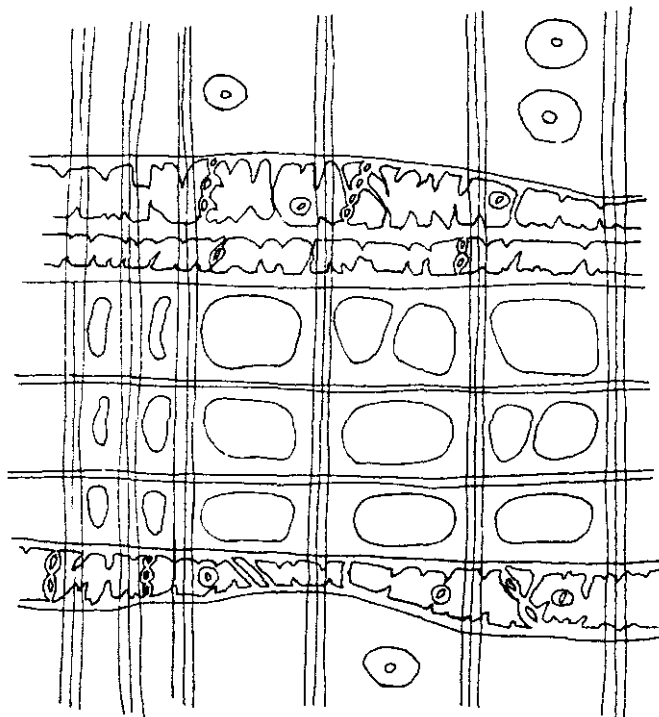


Fig. 2. Left, sketch of radial section of *Pinus resinosa* showing two rows of dentate ray tracheids at the upper margin of the wood ray and a single row on the lower margin; the three center tiers of ray parenchyma illustrate 1-2 large, simple, cross-field pitting. Right, radial section of *Abies magnifica* showing nodular end walls of the ray parenchyma cells in pit-like hollows of the horizontal walls. These hollows are referred to as indentures. A solitary crystal appears in each cell

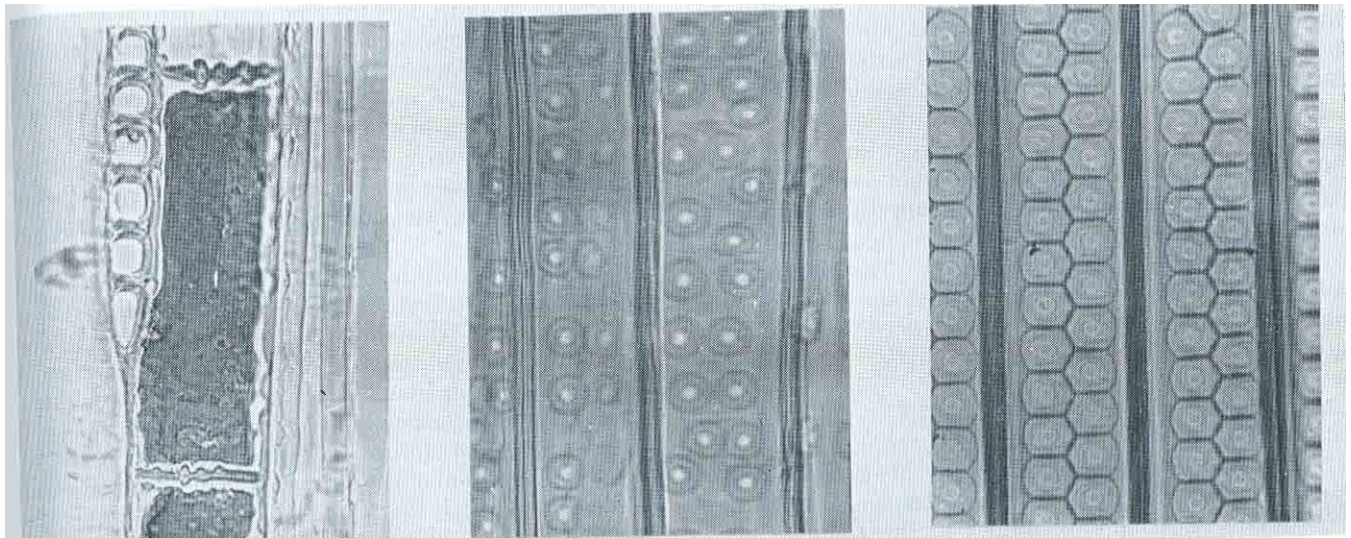


Fig. 3. Left, nodular thickenings of the transverse walls of vertical parenchyma in *Taxodium*; Note dark colored contents. Center, bordered pitting of tracheids multiseriate and opposite in *Sequoia*. Right, bordered pitting multiseriate and alternate in *Agathis*

differentiated from the ray parenchyma by the presence of small bordered pits in the end walls and will be seen there in sectional view.

5. Ray Tracheids Dentate. Ray tracheids exhibiting toothlike projections of the walls are referred to as dentate (Fig. 2, left), and where they appear fused are known as reticulate. Dentate ray tracheids occur in the genus *Pinus* and are associated with those species which show 1-6 pinoid cross-field pitting. They may also occur in combination with 1-2 large, cross-field pitting as in *Pinus resinosa* and *Pinus sylvestris*.

Minute denticulations are regularly present in *Picea* and sporadic in *Larix*. These should be observed from radial sections in the latewood or summerwood portions of the growth rings.

6. Indenture. Depressions at the corners of the ray cells, as observed from radial sections, are referred to as indentures. They appear as very small, pitlike hollows in the horizontal walls in which the ends of the vertical walls stand (Fig. 2, right). These should be observed at a magnification of about 400 times, and used as an identification feature only if they are definitely present. Of our native species, this feature is of greatest diagnostic value in the cedar species.

7. End Walls of Ray Parenchyma Nodular. Bead-like thickenings are sometimes present on the vertical walls of the ray parenchyma cells and can be observed from radial sections. Similar thickenings may occur on the transverse walls of the vertical parenchyma cells (Fig. 2, right).

8. Crystals Present. Crystals appearing in the form of regular prisms, cubes, or octahedra are of common occurrence in one group of the genus *Abies*, and occur

sporadically in *Picea sitchensis* and *Libocedrus decurrens*. The crystals are generally confined to the ray parenchyma cells although they may also be found in the vertical parenchyma in *Abies*. Figure 2, right, illustrates two of the crystal shapes that may occur in *Abies*.

9. Blank.

Cross-field Pitting. The pits occurring on the areas of contact between the ray parenchyma cells and the adjacent vertical tracheids are referred to as cross-field pitting. This may vary from a single large pit that occupies most of the area of the cross-field, as in the case of the white pines, to very small pits that occupy only a small portion of the cross-field areas, as in the case of the spruces. Because the number of pits per cross-field as well as their shape will vary within a given growth ring, it is necessary to limit observations of this feature to the first two or three rows of earlywood or springwood tracheids.

10. Cross-field Pitting, 1-6 Pinoid. Pitting of the 1-6 pinoid type may be without a border or show narrow borders. The individual pits are irregular or variable in shape and size and there may be one to six pits per cross-field. Pitting of this type is limited to the genus *Pinus* and is associated with dentate ray tracheids.

12. Cross-field Pitting, 1-2 (4) Large, Simple. Within this category of 1-2 (4) pits that are large simple, or nearly so are included the large windowlike pits of the white pines, red pine, and the uniformly oval pits of sugar pine (Fig. 2, left). In the case of the white pine and red pine there may be one or two pits per cross-field, whereas in sugar pine there may be two to four pits per cross-field.

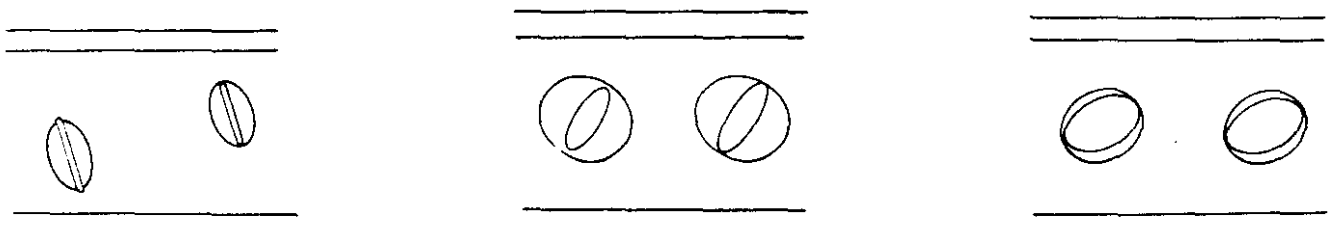


Fig. 4. Cross-field pitting: piceoid (left), cupressoid (center), and taxodioid (right)

12. Cross-field Pitting, Piceoid. Piceoid pits show a narrow and often slightly extended aperture. The aperture in this instance is distinctly narrower than the distance from the aperture to the border (Fig. 4, left).

13. Cross-field Pitting, Cupressoid. Cupressoid pits are those in which the aperture is ovoid and included. The aperture is slightly narrower than or as wide as the space on either side between the aperture and the border (Fig. 4, center).

14. Cross-field Pitting, Taxodioid. Taxodioid pits have a large, ovoid-to-circular included aperture that is wider than the space on either side between the aperture and the border (Fig. 4, right).

15. Blank.

16. Blank.

17. Blank.

18. Bordered Pitting of Tracheids Multiseriate and Opposite. This feature of pitting of tracheids biseriate or triseriate and opposite applies to the common occurrence of 2-4 bordered pits throughout the length of the tracheids and not to occasional groups that may occur at the overlapping portions of the tracheids. This feature is most pronounced in *Sequoia* and *Taxodium* and of infrequent occurrence in other genera; see Fig. 3, center.

19. Bordered Pitting of Tracheids Multiseriate and Alternate. Tracheid pitting may be 2-4 seriate in this group but in alternate arrangement. This feature separates *Araucaria* and *Agathis* from all other conifers; see Fig. 3, right.

20. Spiral Thickenings in Tracheids. Spiral thickenings of tracheids occur only in *Pseudotsuga*, *Taxus*, and *Torreya*. In *Torreya* the thickenings tend to occur in pairs which helps to separate this genus anatomically from *Taxus*. Spiral thickenings should not be confused with the spiral checks that characterize compression wood. Figure 1, right, illustrates spiral thickenings in *Pseudotsuga*.

21. Blank.

22. Vertical Parenchyma Present. When present, parenchyma cells are usually detected without difficulty because of their dark contents. The cells may be limited to the growth ring boundary as in *Abies* and *Tsuga* or may be interspersed throughout the growth ring as in *Taxodium* and *Juniperus*. The dark contents of vertical parenchyma are shown in Fig. 3, left.

23. Transverse Walls of Vertical Parenchyma Nodular. Beadlike thickenings on the transverse walls of vertical parenchyma, similar to those that may occur on the end walls of the ray parenchyma cells, are best observed from tangential sections. The presence of these thickenings and their relative size are very helpful in differentiating between certain species. *Sequoia* and *Taxodium* are readily separated anatomically on this basis: see Fig. 3, left.

24. Blank.

25. Blank.

26. Heartwood Colored. This characteristic is used only with those species that possess a strongly colored heartwood, not where the heartwood exhibits only a slightly darker color than the sapwood. This feature is to be used in the positive sense only.

27. Distinctive Odor. Distinctive odors will range from the rank odor of *Abies lasiocarpa* to the sweetly aromatic odors found in *Thuja plicata* and *Libocedrus*

decurrens. Faint odors may be accentuated by moistening a freshly exposed heartwood surface or by heating a moistened surface. This characteristic is to be used only in the positive sense and to best advantage in those woods in which resin canals are absent.

28. Greasy Feel. A characteristic greasy feel is found in *Taxodium* and in some specimens of *Pinus ponderosa*. This feature is to be used only in the positive sense.

29. Dimpled Grain. Dimpled grain is due to the indentations of the cambium produced by resin cysts in the bark. Thin-barked species such as *Pinus contorta* show this feature prominently on flatsawn surfaces. Dimples also occur frequently in *Pinus ponderosa*, *Picea sitchensis*, and sporadically in several other species.

50. Transition Abrupt. Abrupt transition refers to the sharp definition of the springwood and summerwood of the same growth ring. Figure 5, left, illustrates the abrupt transition in *Larix* and Fig. 5, right, shows the gradual transition of *Pinus strobus*. Species that regularly show an abrupt transition may show a gradual transition in wide growth rings, hence this feature should be used only in the positive sense.

31. Blank.

Geographic Origin

32. Exotic. Species originating outside the boundaries of the continental United States and Canada are known as exotic.

33. Eastern United States. Species whose natural range is east of the Great Plains are listed as from the eastern United States.

34. Western United States. Species whose natural range is west of the Great Plains are classified as from the western United States.

USE OF DATA TABLE AND CARD KEY IN IDENTIFICATION

The data in Table I may be used independently as a key or the data may be transposed to cards and the identification may be accomplished by sorting. The suggested layout of the card key is shown in Fig. 6.

Use of Tabulated Data as a Key

The plus marks in Table I represent the presence of

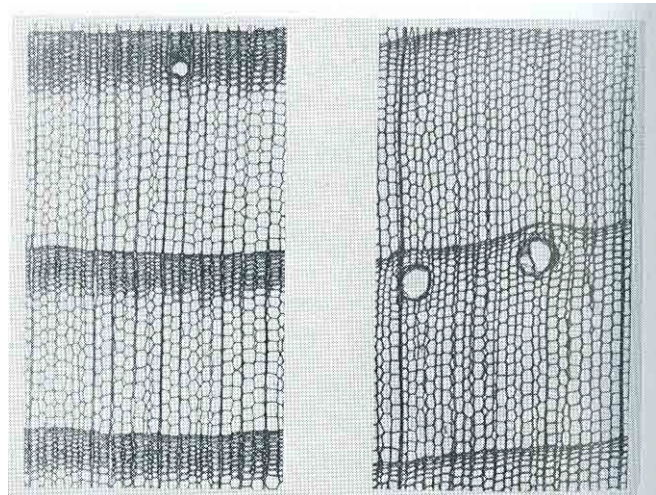


Fig. 5. Left, abrupt transition in *Larix* and one small vertical resin canal in the summerwood. Right, gradual transition in *Pinus strobus* and two large vertical resin canals at the margin of the summerwood

Table 1. Identification Chart and Data for Preparation of Card Keys

Diagnostic features	Feature number	Resin canals present										Resin canals absent																										
		Epithelium thick-walled					Epithelium thin-walled					Ray tracheids present					Ray tracheids absent																					
												End walls of ray cells nodular					End walls of ray cells smooth																					
		Pseudotsuga	Larix laricina	Larix occidentalis	Picea sitchensis	Picea spp.	Picea spp.	Pinus spp.	Pinus spp.	Pinus resinosa	Pinus strobus	Pinus monticola-lambertiana	Pinus spp. (sectant)	Pinus spp. (penyon)	Chamaecyparis nootkatensis	Tsuga heterophylla	Tsuga canadensis	Abies balsamea	Abies spp. (white fir)	Abies lasiocarpa	Juniperus spp.	Juniperus spp.	Fitseria cupressoides	Libocedrus decurrens	Cupressus arizonica	Taxus brevifolia	Torreya taxifolia	Thuja plicata	Thuja occidentalis	Cupressus lusitanica	Chamaecyparis lawsoniana	Chamaecyparis thyoides	Taxodium distichum	Sequoia sempervirens	Araucaria-Agathis			
Canals																																						
Present	1	+	+	+	+	+	+	+	+	+	+	+	+																									
Epithelium thick-walled	2	+	+	+	+	+	+																															
	3																																					
Rays																																						
Tracheids present	4	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+																						
Tracheids dentate	5						+	+	+																													
Indenture	6	+	+	+	+	+										+	+	+	+	+	+	+	+	+	+													
End walls nodular	7	+	+	+	+	+					+	+	+	+	+	+	+	+	+	+	+	+	+	+	+			+	+									
Crystals	8																																					
	9																																					
Cross-field pitting																																						
1-6 pinoid	10						+	+																														
1-2 (4) large	11								+	+	+																											
Piceoid	12	+	+	+	+	+							+	+		+																						
Cupressoid	13														+						+	+	+	+	+	+	+	+										
Taxodioid	14																	+	+	+																		
	15																																					
	16																																					
	17																																					
Tracheids																																						
Pits 2-4 opposite	18		+	+	+											+																						
Pits 2-4 alternate	19																																					
Spirals	20	+																																				
	21																																					
Vertical parenchyma																																						
Present	22	+													+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+		
Transverse walls nodular	23	+													+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+		
	24																																					
	25																																					
General																																						
Heartwood	26	+	+	+	+			+	+	+	+	+	+																									
Odor	27	+													+					+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+		
Greasy	28																																					
Dimpled	29																																					
Transition abrupt	30	+	+	+				+	+	+						+																						
	31																																					
Origin																																						
Exotic	32																							+														
Eastern	33		+			+		+	+	+																												
Western	34	+		+	+		+	+	+		+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	

the given characteristic for a particular species or group of species. Blank spaces indicate the absence of the feature. The data are so tabulated that the species and groups are reduced to a small number within each category, which permits easier comparisons leading to identification. Under the heading "Resin canals present" and the subheading "Epithelium thin-walled" we find all the pines. Further separation may be accomplished by means of the presence or absence of dentate ray tracheids and still further by means of the cross-field pitting. This elimination may be continued until the identification is pinpointed to a single group.

In transposing the data from Table I to marginally perforated cards, the features marked with a plus (+) are to be indicated on the card by a notch. Features which are shown as blanks in the table remain unnotched on the card. For example, perforation no. 1 is designated as "Resin canals present" and, if notched, indicates the presence of this feature; if left unnotched, this indicates the absence of resin canals. Upon inserting a needle or thin rod into a pack of cards, those which are notched at a particular perforation will fall from the pack when the entire series is lifted and gently shaken. This process is repeated for a succession of the more important characters of the sample until a single card falls from the pack and the determination is accomplished.

When the region of growth is positively known, it is usually desirable to start sorting at this point; in so doing the card pack will be reduced by about one half and permit more accurate diagnosis.

Preferred sequence of operation for woods with resin canals:

Region of growth if positively known.

Resin canals present or absent—cards falling from pack are those woods in which canals are present.

Sort for thick-walled epithelium—cards falling from pack are woods with thick-walled epithelium and those remaining on needle are those with thin-walled epithelium.

If epithelium is thin-walled, then sort for ray tracheids dentate—those falling from the pack have dentate ray tracheids and those remaining on the needle have smooth or non-dentate ray tracheids.

Next sorting should be in cross-field pitting, followed by any other feature which is presented by the particular sample.

Preferred sequence of operation for woods without resin canals:

Region of growth if positively known.

Resin canals present—the cards remaining on the needle are the woods without resin canals.

Ray tracheids common—cards falling from pack are those in which ray tracheids are common and those remaining on needle are without ray tracheids.

End walls of ray cells nodular—cards falling from pack contain this feature and cards remaining on needle are those woods in which the end walls are smooth.

Continue with type of cross-field pitting, vertical parenchyma present, vertical parenchyma if present with or without nodular transverse walls, bordered pitting 2-4 seriate and whether alternate or opposite, indenture, followed by any other feature pertinent to the specimen.

Species having highly distinctive characteristics can be sorted very quickly without going through the above operations. Douglas-fir, for example, can be arrived at in two sorts, the first for spiral thickenings present in the vertical tracheids and the seed for resin canals present. The red pines can likewise be removed

18	2-4 opposite	Tracheids	Resin canals	1
19	2-4 alternate	Tracheids	Epith. thick	2
20	Spirals	Tracheids	Canals	3
21			Ray tracheids	4
22	Parenchyma	Parenchyma	Dentate	5
23	Nodular	Parenchyma	Indenture	6
24			Walls nodular	7
25			Crystals	8
26	Heartwood			9
27	Odor	General	1-6 Pinoid	10
28	Greasy	General	1-2(4) Large	11
29	Dimpled		Piceoid	12
30	Trans. abrupt		Cupressoid	13
31			Taxodioid	14
32	Exotic	Origin		15
33	Eastern	Origin		16
34	Western	Origin		17

Fig. 6. Suggested layout of features for card sorting key

in two sorts—foray tracheids dentate, and cross-field pitting 1-2(4) large. A single sort at tracheid pitting multiseriate and alternate will immediately bring out the Araucariaceae (Agathis and Araucaria).

SUPPLEMENTARY NOTES

Summarized here are the important anatomical features of the various genera of conifers contained in the key and supplementing the data in Table I. Means of distinguishing the individual species are given here because such specific identifications often depend on characteristics that are insufficiently precise for inclusion in the list of key features. The distinctive features of each genus and the degree to which the genus may be subdivided in anatomical or other features are also noted.

Abies

Nine species of *Abies* are native to the United States and Canada. The only eastern species of commercial importance is balsam fir (*A. balsamea* (L.) Mill.). The other eastern species, Fraser fir (*A. fraseri* (Pursh.) Poir.) is extremely limited in its distribution and hence of little importance. The western species, Pacific silver fir (*A. amabilis* (Dougl.) Forbes), white fir (*A. concolor* (Gord. & Glend.) Lindl.), grand fir (*A. grandis* (Dougl.) Lindl.), subalpine fir (*A. lasiocarpa* (Hook.) Nutt.), California red fir (*A. magnifica* A. Murr.), and noble fir (*A. procera* Rehd.) are of considerable importance from the production standpoint. The bristlecone fir (*A. bracteata* D. Don) is limited to Monterey County, Calif., and does not enter commercial channels. The latter species may be readily recognized by the very frequent occurrence of traumatic resin canals.

Woods belonging to this genus may be readily recognized anatomically by the following combination of features: Resin canals lacking, ray tracheids lacking, end walls of ray parenchyma nodular, and cross-field pitting taxodioid.

The woods of this genus may be further subdivided into two groups on the basis of the color of the ray cell contents. In the eastern species and in subalpine fir the ray cell contents are colorless or at most a very pale yellowish color, and frequently tend to form a reticulum within the cells so that detection of the end walls may be somewhat difficult. These three species are further characterized by the lack of crystals in the ray parenchyma. The wood of *Abies lasiocarpa* frequently has a rather foul odor and the knots are distinctly yellowish in color.

The remaining species, all western, have ray cell contents that are distinctly reddish brown. Crystals may be found in all species of this series but are common to abundant in *A. magnifica* and *A. concolor*, and sparse or infrequent in *A. amabilis*, *procera*, and *grandis*. For practical considerations, the western species are classed collectively as "white fir."

Araucariaceae

In the *Araucariaceae* family are three species that enter commercial channels: Brazilian araucaria or "Parana pine" (*Araucaria angustifolia* (Bert.) O. Ktze.) of Brazil; klinki pine (*Araucaria klinkii* Lauterb.) of Borneo; and almáciga or "sakar" (*Agathis philippinensis* Warb.) of the Philippine Islands. The group as a whole is easily separated from all other coniferous woods by the 2-4 seriate bordered pitting of the vertical tracheids which is in alternate arrangement (Fig. 5, right). The two *Araucarias* are separated from *Agathis* by the presence of leaf traces in the *Araucarias*. These traces appear as tiny pin knots arranged obliquely as seen on the surface of flatsawn boards. Brazilian araucaria tends to be yellowish in color and frequently shows reddish or pinkish streaks while klinki pine is pinkish in color, is lighter in weight, and does not show reddish streaks. In color and luster, klinki pine is very similar to Sitka spruce (*Picea sitchensis* (Bond.) Carr.).

The lack of growth rings in the family also helps to separate these imported woods from native softwood species.

Chamaecyparis

Three species of *Chamaecyparis* occur within the United States and Canada: Atlantic white-cedar (*C. thyoides* (L.) B.S.P.), of the coastal plain from southern Maine to Mississippi; Port-Orford-cedar (*C. lawsoniana* (A. Murr.) Parl.) of southwestern Oregon and northwestern California; and Alaska-cedar (*C. nootkatensis* (D. Don) Spach), which ranges from southeastern Alaska to northwestern California.

The three species vary appreciably and possess no features in common that would distinguish the genus. The heartwood of *C. nootkatensis* is yellowish and that of *C. lawsoniana* ranges from yellowish-white to a pinkish-brown or very pale brown. In both species there is no sharp contrast in color between heartwood and sapwood. The heartwood of *C. thyoides* is a light brown with a pinkish or reddish cast and is sharply demarcated from the sapwood.

From the standpoint of odor the three species differ rather markedly. The heartwood of *C. thyoides* is aromatic and resembles that of northern white-cedar (*Thuja occidentalis* L.); the heartwood of *C. lawsoniana* is strongly aromatic with a pungent, gingerlike odor; and the heartwood of *C. nootkatensis* has an odor that resembles that of raw potatoes.

Anatomically they may be separated as follows: *C. nootkatensis* has ray tracheids that are quite large and frequently comprise the entire wood ray; the end walls of the vertical parenchyma are nodular, as are also the end walls of the ray parenchyma cells; and indenture is present. *C. lawsoniana* and *C. thyoides* are similar in that they lack ray tracheids and the end walls of the ray cells are smooth and without indenture. They differ in that the transverse walls of the vertical parenchyma are nodular in *C. lawsoniana* and smooth in *C. thyoides*.

Cupressus

Of the various species of *Cupressus*, only two are occasionally encountered: Arizona cypress (*C. arizonica* Greene) of the southwestern United States and Bentham cypress (*C. lusitanica* Miller) of Mexico and Guatemala. The heartwood of *C. arizonica* is not differentiated from the sapwood, is yellowish in color and aromatic. The heartwood of *C. lusitanica* is a pale reddish brown in color and is distinctly separated from the light-colored sapwood. The heartwood of *C. lusitanica* is also aromatic.

Anatomically *C. arizonica* has many of the features common to *Juniperus*; however, *C. arizonica* tends to have lower rays and the end walls of the ray parenchyma cells are not as conspicuously nodular as in *Juniperus*. When heartwood is present, the two genera can be readily separated.

C. lusitanica differs from *C. arizonica* in that the end walls of the ray cells are not nodular in *C. lusitanica* and indenture is lacking or obscure.

Fitzroya

The single species of *Fitzroya*, alerce (*F. cupressoides* (Mol.) Johnston) is native to southern Chile. Superficially the wood resembles that of redwood (*Sequoia sempervirens* (D. Don) Endl.) but may be differentiated when split radial faces of the two species are compared. In redwood the rays are two to three times higher than those of alerce. Anatomically, alerce may be separated from redwood by the presence of indenture, the end walls of the ray cells are nodular, cupressoid pitting is present, and the wood rays are usually under 12 cells in height.

Juniperus

The heartwood of the various species of *Juniperus* is readily distinguishable from all other conifers by its distinctive odor that is generally classified as "cedar-chest" or "pencil" odor. On the basis of heartwood color, two groups may be differentiated: (1) those with a deep-red or purplish-red color as in eastern redcedar (*J. virginiana* L.), southern redcedar (*J. silicicola* (Small) Bailey), and Rocky Mountain juniper (*J. scopulorum* Sarg.) and (2) those with a brownish-colored heartwood that includes all of the western species except *J. scopulorum*.

Anatomically; *Juniperus* may be distinguished from other woods with a highly colored and aromatic heartwood by its abundant vertical parenchyma with nodular transverse walls, ray parenchyma with abundantly nodular ends walls, indenture, dark-colored ray cell contents, and intercellular spaces between the vertical tracheids.

Larix

Three species of *Larix* occur in the United States and Canada: Tamarack (*L. laricina* (Du Roi) K. Koch) of the northeastern United States and from southeastern Canada to the Yukon; western larch (*L. occidentalis* Nutt.) of the northwestern United States and southeastern British Columbia; and subalpine larch (*L. lyallii* Parl.), which is a high-elevation species having the same range as *L. occidentalis* but not of commercial importance.

On the basis of anatomical structure, *Larix* is readily separated from other genera that have resin canals with thick-walled epithelium cells. This separation is made on the basis of the abrupt transition from earlywood to latewood and the bordered pitting of the vertical tracheids, which is frequently biseriate. Western larch may be separated from tamarack using the following combination of features: Heartwood of *L. occidentalis* is russet brown, whereas in *L. laricina* it is a yellowish brown; latewood is generally wider and more pronounced in *L. laricina* than in *L. occidentalis*; growth rings are generally more uniform in *L. occidentalis* than in *L. laricina* and bordered pitting is more commonly biseriate in *L. occidentalis* than in *L. laricina*. The locality of growth affords a very reliable means of separation of these two species.

Libocedrus

The single American species of *Libocedrus*, incense-cedar (*L. decurrens* Torr.), occurs from western Oregon to southern California.

The heartwood is reddish brown to dull brown and sometimes has a purplish cast. The odor of the heartwood is distinctly aromatic and rather similar to that of western redcedar (*Thuja plicata* Donn). Anatomically, it may be distinguished from other genera that have nodular thickenings on both the end walls of the ray cells and the transverse walls of the vertical parenchyma since *Libocedrus* lacks indenture which is present in the other genera.

Picea

Seven species of *Picea* occur in the United States and Canada, although only five are of commercial importance. The most important species are white spruce (*P. glauca* (Moench) Voss) and black spruce (*P. mariana* (Mill.) B.S.P.), which range from the northeastern United States across Canada to Alaska; red spruce (*Picea rubens* Sarg.) which ranges from the Middle Atlantic States to Nova Scotia; Engelmann spruce (*P. engelmannii* Parry) of the Rocky Mountain region; and Sitka spruce (*P. sitchensis* (Bong.) Carr.), which ranges from southern Alaska to northeastern California. Brewer spruce (*P. breweriana* S. Wats.) of southwestern Oregon and northern California and blue spruce (*P. pungens* Engelm.) of the Rocky Mountain region do not enter commercial markets.

The heartwood of *P. sitchensis* is pinkish in color and is sharply demarcated from the white sapwood: The heartwood of the other species is light colored and there is no difference between heartwood and sapwood. Anatomically *P. sitchensis* can be separated from the other species on the basis of the abundant yellowish-brown deposits in the ray cells and the frequent occurrence of biseriate bordered pitting of the vertical tracheids. The other species only show occasional pale yellowish-colored deposits and the tracheid pitting is uniseriate. *P. sitchensis* occasionally has crystals in the ray cells, which is a further aid in separating this wood from the other spruces. Split tangential faces of *P. sitchensis* commonly show dimpled grain. Separation of the other species is not practical except in some instances where the geographical source is known.

Anatomically, *Larix* and *Picea* are very similar; however, separation may be accomplished by the abrupt transition from earlywood to latewood in *Larix* and gradual in *Picea*, and by the presence of minute denticulations on the walls of the ray tracheids in *Picea* but rare in *Larix*.

Pinus

The members of the genus *Pinus* are readily separated from all other conifers by their resin canals, which have thin-walled epithelium cells. Specific identification is generally not possible, however, the genus can be subdivided into five groups on the basis of the ray tracheids and the type of cross-field pitting.

Ray tracheids nondentate.

Cross-field pitting 1-2large, simple or nearly so

Eastern white pine (*P. strobus* L.)

Western white pine (*P. monticola* Dougl.)

Included here are the stone pines of small economic importance which are: whitebark pine (*P. albicaulis* Engelm.) and limber pine (*P. flexilis* James).

Cross-field pitting 2-4 ovoid.

Sugar pine (*P. lambertiana* Dougl.)

Cross-field pitting piceoid.

Species of little importance and comprising two groups:

Foxtail pines: bristlecone pine (*P. aristata* Engelm.) and

foxtail pine (*P. balfouriana* Grev. & Balf.)

Pinyon pines: Mexican pinyon (*P. cembroides* Zucc.);

pinyon (*P. edulis* Engelm.); singleleaf pinyon (*P.*

monophylla Torr. & Frem.); and Parry pinyon (*P. quadrifolia* Parl.)

Raytracheids dentate to reticulate.

Cross-field pitting 1-2large, simple or nearly so.

Red pine (*P. resinosa* Ait.) and the European Scotch pine (*P. sylvestris* L.)

Cross-field pitting 1-6pinoid.

Western species, principally ponderosa pine (*P. ponderosa*

Laws.) Jeffrey pine (*P. jeffreyi* Grev. & Balf.), and

lodgepole pine (*P. contorta* Dougl.)

Eastern and southern species, principally jack pine (*P.*

banksiana Lamb.); slash pine (*P. elliotii* Engelm.);

longleaf pine (*P. palustris* Mill.); shortleaf pine (*P.*

echinata Mill.) and loblolly pine (*P. taeda* L.).

In this group characterized by 1-6 pinoid pitting, separation of the eastern and western species may generally be made by the more prominent latewood found in the eastern species and particularly in the more important southern species. *P. elliotii* and *P. palustris* frequently show multiple latewood bands within individual growth rings which aids in separating these two species from all the others. *P. palustris* can also be separated from the other eastern and southern species when pith is present in the specimen. In this species

the pith is commonly 0.2 in. in diameter while in the other species it is 0.1 in. in diameter or smaller.

Dimpled grain is distinctive of *P. contorta* and occurs rather frequently in *P. ponderosa*. In the latter species, however, the dimples are larger and fewer per unit area than those of *P. contorta*.

Pseudotsuga

The Douglas-fir of commerce is the produce of *Pseudotsuga menziesii* (Mirb.) Franco, occurring from southwestern British Columbia southward through California, and *P. menziesii* var. *glauca* (Beissn.) Franco, which is native to the Rocky Mountain region.

Another species, bigcone Douglas-fir (*P. macrocarpa* (Vasey) Mayr) is limited to the mountains of southern California and is not commercially important.

Pseudotsuga is readily distinguished from all other native conifers by the combination of resin canals present and the occurrence of spiral thickenings of the vertical tracheids.

The heartwood of Douglas-fir is reddish or pinkish and occasionally may have a russet-brown color. The odor of the heartwood is highly distinctive for this species but difficult to describe.

Sequoia

Only one species of Sequoia, Redwood (*S. sempervirens* (D. Don) Engl.), is of commercial importance. The other species, giant sequoia (*S. gigantea* (Lindl.) Decne.), occurs in protected groves. Both species are native to California.

The light red to deep reddish-brown heartwood readily separates redwood from other conifers with colored heartwood. The wood is without odor.

Anatomically, it is identified by its biseriate or triseriate bordered pitting, end walls of ray cells without nodular thickenings, taxodioid cross-field pitting, and relatively inconspicuous nodular thickenings on the transverse walls of the vertical parenchyma. Anatomically it is very similar to *Taxodium* which differs from Sequoia in that the transverse walls of the vertical parenchyma are conspicuously nodular. (See Fitzroya for difference between that genus and Sequoia.)

Taxodium

Baldcypress (*Taxodium distichum* (L.) Rich.) and the variety *nutans* (Ait.) Sweet supply the cypress of commerce. *T. distichum* occurs in the coastal plain from southern Delaware to southeastern Texas and in the Mississippi River valley as far north as Missouri and southern Illinois. The variety *nutans* occurs in the coastal plain from southeastern Virginia to southeastern Louisiana.

The heartwood of *Taxodium* is quite variable in color and ranges from yellowish to dark brown, reddish brown, or almost black. The heartwood is further characterized by its greasy or oily feel and often by a rancid odor.

Anatomically it is very similar to Sequoia but has more conspicuous nodular thickenings on the transverse walls of the vertical parenchyma. The cross-field pitting of *Taxodium* may be taxodioid or cupressoid.

Taxus

Two species of *Taxus*, Florida yew (*T. floridana*

Nutt.) and Pacific yew (*T. brevifolia* Nutt.), are native to the continental United States; however, only the Pacific yew enters the market. *T. floridana* is rare and local in Gadsden and Liberty Counties of northwestern Florida. *T. brevifolia* occurs from southeastern Alaska to northern California and in the northern Rocky Mountain region.

The heartwood ranges in color from bright orange to russet brown and is without odor. Anatomically it can be separated from all other American conifers except *Torreya* by its spiral thickenings in the vertical tracheids and lack of resin canals. In *Taxus* the cross-field pits are 8 to 10 μ in diameter while in *Torreya* they are about 6 μ in diameter (see *Torreya*).

Thuja

Two species of *Thuja* occur in the United States and Canada. Northern white-cedar (*T. occidentalis* L.) is found in southeastern Canada and northeastern United States to the Lake States region, while Western redcedar (*T. plicata* Donn) ranges from southeastern Alaska to northwestern California and also in the northern Rocky Mountain region.

The heartwood of *T. occidentalis* is a pale brown with a faint but characteristic cedary odor; the heartwood of *T. plicata* is reddish or pinkish brown to dull brown with a much stronger and spicy aromatic odor.

Anatomically the genus is defined by taxodioid cross-field pitting, end walls of ray cells smooth, indentures distinct, transverse walls of vertical parenchyma nodular. *T. occidentalis* may be separated anatomically from *T. plicata* by the more frequent biseriate bordered pitting and the more abundant parenchyma of *T. plicata*. The end walls of the ray cells may occasionally show nodular thickening, particularly near the growth ring boundaries. In *T. occidentalis* the ray cell contents appear reticulate and tend to obscure the end walls, while in *T. plicata* the contents of the ray cells are more uniformly distributed and do not obscure the end walls.

Torreya

The genus *Torreya* consists of two rather rare species: Florida torreyia (*T. taxifolia* Am.) of southwestern Georgia and northwestern Florida and California torreyia (*T. californica* Torr.) of central California.

The heartwood is a clear yellow color and has a rather distinctive and somewhat unpleasant odor.

Anatomically *Torreya* is very similar to *Taxus*, however, it may be separated from *Taxus* by the paired appearance of the spiral thickenings, and the smaller (about 6 μ) size of the cross-field pits.

Tsuga

The genus *Tsuga* consists of four species: western hemlock (*T. heterophylla* (Raf.) Sarg.) and mountain hemlock (*T. mertensiana* (Bong.) Carr.) of the northwestern United States and southwestern Canada, and eastern hemlock (*T. canadensis* (L.) Carr.) and Carolina hemlock (*T. caroliniana* Engelm.) of the eastern region. *T. canadensis* occurs in the Lake States, northeastern United States, and southeastern Canada, and *T. caroliniana* is of limited occurrence in the Central Appalachians.

The woods of this genus show no difference between

sapwood, and heartwood and bear a close resemblance to the wood of the western *Abies* with which they are frequently confused. Anatomically they are readily separated from the *Abies* by the ray tracheids which are always present in *Tsuga* and rarely present in *Abies*. The eastern species may be separated from the western species by the more abrupt transition from earlywood to latewood and the frequent biseriate bordered pitting that occurs in the former species. The western *Tsuga* may be separated from the western *Abies* by its more

uniform and somewhat purplish color as seen on smooth cut end-grain surfaces and finer texture as seen with a hand lens. In the *Abies* species the color change from earlywood to latewood is more abrupt and the summerwood generally is light brown. The texture of the western *Abies* is distinctly coarser than that of *Tsuga heterophylla*.

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